Biochemical Alterations after Single Oral Dose of Monocrotophos in *Bubalus bubalis*

Harpal S. Sandhu and Jitendra K. Malik¹

Department of Pharmacology, College of Veterinary Science, Punjab Agricultural University, Ludhiana-141004, India

The wide spread use and in many cases their presence and/or concentration in food chain, organophosphorus insecticides constitute a major group of potential health hazards to man and livestock. Like other organophosphorus insecticides, the biological activity of monocrotophos is not limited to insects only but it is also toxic to mammals and birds (Stickel 1975. Mendelssohn 1977). The indiscriminate and inadvertent use of monocrotophos poses health hazards to domestic animals and birds exposed either by ingestion of insecticide sprayed fodder or inhalation from the contaminated environment (Shlosberg et al. 1980). The detailed information on the acute toxicity of monocrotophos is, however, absolutely lacking in buffalo Previous reports have indicated that species. susceptibility of buffaloes to organophosphorus insecticides differ markedly from other species of animals including rodents (Umetsu et al. 1977, Clarke et al. 1981). The present study was therefore undertaken to investigate the effects of single oral administration of monocrotophos on plasma cholinesterase (ChE), serum aminotransferases and total plasma proteins.

MATERIALS AND METHODS

Twelve healthy male buffalo calves (70-130 kg), purchased from local market, were maintained on green fodder (maize or barseem) and wheat straw. Water was provided ad libitum. The calves were randomly divided into three groups, four animals each. Based on preliminary trials, animals of these groups received single dose of monocrotophos (Nuvacron, 36 EC w/w, Hindustan Ciba-Geigy Ltd., Bombay) at the dose rate of 10, 20 and 40 mg/kg body weight. The requisite quantity

Send reprint requests to Dr.Harpal S. Sandhu at the above address

¹ Present address: Professor-cum-Head, Department of Pharmacology, Gujarat Agricultural University, Anand, India

monocrotophos was suspended in 100 ml of tap water and given by drenching. All the calves were fasted for 16 hours prior to administration of insecticide.

Monocrotophos treated animals were observed for the appearance of any toxic symptoms and extent of lethality. The surviving animals were kept under close observation over a period of 28 days after the ingestion of insecticide. Blood samples were collected by jugular venipuncture prior to and at different time intervals after monocrotophos administration. Plasma and serum were separated at room temperature. Plasma ChE (E.C. 3.1.1.8) was determined according to the methods of Voss (1970) as modified by Morai et al. (1976). The levels of serum aspartate aminotransferase (E.C. 2.6.1.1); serum alanine aminotransferase (E.C. 2.6.1.2) and plasma protein were measured by the procedures of Wotton (1964). The statistical significance between two means based on individual observations was assayed by student's t-test.

RESULTS AND DISCUSSION

Monocrotophos was administered in single doses of 10, 20 and 40 mg/kg body weight by oral route. At all doses it produced mild to moderate degree of toxic symptoms which were characteristics of typical anticholinesterase poisoning. The severity of toxic symptoms were dose dependent. The lowest dose (10 mg/kg) was not lethal whereas 20 and 40 mg/kg doses produced 50 and 100 per cent lethality within 106-116 and 14-32 h, respectively. The occurance of toxic symptoms following monocrotophos administration was considerably delayed. The delay in appearance of toxic symptoms may be due to slower absorption of monocrotophos from the gastrointestinal tract; as absorption of xenobiotics from the qastrointestinal tract is a complex process invol ving interaction of many factors i.e. lipophilicity, binding, pH, degree of ionization etc. (Hwang and Schanker 1974; Ahdaya et al. 1981). In buffalo species an other organophosphorus insecticide, dimethoate. has also shown delayed toxicity following acute poisoning (Singh 1981).

All the doses of monocrotophos produced marked inactivation of plasma cholinesterase. The extent of inhibition of plasma cholinesterase correlated well with the severity of toxicity in buffalo calves given 20 and 40 mg/kg doses of monocrotophos. Maximal inactivation of plasma cholinesterase was observed at 24 h and at this time animals showed severe toxic symptoms. However no relationship between recovery of apparent toxic symptoms and reactivation of plasma cholinesterase enzyme could be established in animals survived

Table 1. Effect of single oral administration of monocrotophos on plasma cholinesterase and total plasma proteins in buffalo calves

Dase			Time	ıfter admin	Time after administration (h)	h)		
(mg/kg) 0	0	2	7	12	24	48	168 672	672
10	175.2± 2.1	Plasma cho 147.4±**	lasma cholinesterase (n mol acetylthiocholine hydrolysed/min/ml) $47.4\pm **$ $137.1\pm **$ $112.6\pm **$ $105.6\pm **$ $114.3\pm **$ $150.7\pm **$ 2.2 4.2 6.4 8.4 11.0	(n mol ace 112.6±**	tylthiochol 105.6±** 8.4	ine hydroly 114.3±** 8.4	sed/min/ml) 150.7±**	170.8±
20	176.8± 1.0	146.9±** 5.7	130.0±** 7.9	66.9±**	52.2±** 2.9	45.2±** 4.5	71.8 ^a	162.1 ^a
40	178.5± 5.0	133.8±** 5.7	102.3±** 9.2	32.1±** 2.9	32.6 ^b			
		Total plas	Total plasma proteins	(g/100 ml)				
10	7.0± 0.0	7.5±* 0.1	7.6±**	8.1±** 0.1	8.4±** 0.1	8.4±** 0.2	7.2± 0.1	7.8± 0.0
20	7.2± 0.1	7.9±* 0.2	8.2±** 0.1	9.0 _± **	9.0±** 0.4	8.4±** 0.4	7.4ª	2.0°
07	7.4±	8.9±* 0.2	9.6±**	10.6±**	11.0 ^b			

Values given are mean \pm SE of the results from 4 animals unless otherwise stated. a - Value is mean of 2 animals b - Value of 1 animal * P / 0.05 * P / 0.01

Table 2. Effect of single oral administration of monocrotophos on serum aminotransferases in buffalo calves

(mg/kg) 0	0	2	7	12	54	8 7	168	672
	Serum	ırtate	aminotransferase (n mol	rase (n mol	pyruvate f	pyruvate formed/min/ml		
10	59.7± 64 1.0 C	64.8± 0.8	66.2±*	75.4±**	78.7±** 2.2	76.5±** 3.2	63.9± 2.7	58.6±
20	63.4± 0.5	72.3±** 0.5	75.9±** 0.8	90.2±** 2.0	94.4±** 2.1	90.5±** 2.4	65.3a	60.3 ^a
40	65.3±	83.8±**	96.9 _± **	137.1±**	140.7 ^b			
	Serum	al	anine aminotransferase (n mol pyruvate formed/min/ml)	se (n mol p)	ruvate for	med/min/ml)		
10	50.4± 1.1	60.4±* 1.9	62.6±**	82.5±** 5.7	83.7±**	77.6±** 2.9	56.5年2.3	50.9±
20	46.6± 1.8	69.8±**	80.4±** 2.8	108.6±** 3.7	115.3±** 6.8	113.1±** 4.8	69.8 _a	44.3ª
40	50.4±	107.5±**	119.7±** 2.7	174.0±** 4.9	192.9 ^b			

Values given are mean \pm SE of the results from 4 animals unless otherwise stated. a - Value is mean of 2 animals. b - Value of 1 animal \star P/ 0.05

**P/0.01

after monocrotophos administration as toxic symptoms in surviving animals were recovered within 24-31 h but during this time plasma cholinesterase activity remained markedly inhibited (P / 0.01). Similar results have been reported with various other organophosphorus insecticides in different species of animals (Jovic 1974, Malik et al. 1978).

There was significant elevation in the serum levels of aspartate and alanine aminotransferases (Table 2). At 24 h the levels of aspartate and alanine aminotransferases were increased to the extent of 32.4-115.4 and 65.9-282.4 per cent, respectively. The increase levels of serum aminotransferases have been used as indicator of cellular damage (Drotman and Lawhorn 1978), increased plasma membrane permeability (Ramazzotto and Carlin 1978) or altered metabolism of these enzymes (Dinman et al. 1963). The oral administration of organophosphorus esters is known to severely effect the liver, an organ primarily involved in their activation and/or detoxification (Murphy 1980). In addition, organophosphorus compounds increases the permeability of artificially prepared lipid membranes (Antunes-Madeira and Madeira 1979). From the present results, however, it could not be ascertained whether monocrotophos mediated elevation of these enzymes was a result of increased plasma membrane permeability or cellular damage or a combination of these factors.

The oral administration of monocrotophos produced marked elevation in the levels of plasma proteins (Table 1). It has been suggested that organophosphorus insecticides may stimulate the growth of cellular proteins and RNA in animals (Puga et al. 1974). Similar to our findings Mihai et al (1972) also observed increased levels of aminoacids in blood and liver of horses given trichlorfon.

REFERENCES

Ahdaya SM, Monroe JE, Guthrie FE (1981) Absorption and distribution of intubated insecticides in fasted mice. Pest Biochem Physiol 16: 38-46

Antunes-Madeira MC, Madeira VMC (1979) Interaction of insecticides with lipid membranes. Biochem Biophys Acta 550: 384-392

Clarke ML, Harvey DG, Humphreys DJ (1981) Veterinary toxicology. 2nd edn English Language Book Soc and Bailliere Tindal, London pp 146-156

Dinman BD, Handi EA, Fox CF, Frajola WJ (1963) Carbon tetrachloride toxicity. Hepatostructural and enzymatic changes. Arch Environ Health 7: 630-646

- Drotman RB, Lawhorn GT (1978) Serum enzymes as indicatiors of chemically induced liver damage. Drug Chem Toxicol 1: 163-171.
- Hwang SW, Schamuker LS (1974) Absorption of carbaryl from the lung and small intestine of the rat Environ Res 7: 206.
- Jovic RC (1974) Correlation between signs of toxicity and some biochemical changes in rats poisoned by soman. European J Pharmac 25: 159-164
- Malik JK, Verma SP, Garg BD, Ahmad A (1978) Studies on blood enzymes during acute toxicity of 0-ethyl-S, diphenyl phosphorodithioate (Hinosan) in buffalo calves. Indian J Pharmac 10: 7-14.
- Mendelssohn H (1977) Mass mortality of birds of prey caused by Azodrin an organophosphorus insecticide. Biological Censervation 11: 163-170.
- Mihai M, Badescu C, Lorgulescu PA, Harisanidi S (1972) Changes in the tissue proteins of animals (guinea pig and horses) given pesticides (trichlorfon and Lindane). Zooprofilassi 27: 83-90.
- Morai K, Usbiyama S, Satoh T, Kuga T (1976) Enzyme induction of repeated administration of tetrachlorvinphos in rats. Toxicol Appl Pharmacol 37: 377-386.
- Murphy SD (1980) Pesticides In: Doull J, Klassen CB, Amdur MD (eds) Toxicology: The basic science of poisons. Macmillan, New York pp 398-400.
- Puga FR, Rodrigues Lark MA (1974) Effect of dimethioate and propoxur on the metabolism of IB-RS-2 cells and on their susceptibility to foot and mouth disease.

 Arquinos do Instituto Biologic Sao-Paulo, Brazil 3: 141-145
- Ramazzotto LJ, Carlin R (1978) Effects of DMSO on SGOT during hypothermia in adrenalectomized rats. Life Sci 22: 329-336.
- Shlosberg A, Egyed MN, Hanji V (1980) Monocrotophos poisoning in geese caused by drift from crop spraying Refuan Veterinariah 37: 42-44
- Singh SD (1981) Toxicological and biochemical studies of O, O-dimethyl S-(N-ethyl carbamoyl methyl) phosphorodithioate (Dimethoate) in Bubalus bubalis. Ph.D Thesis Punjab Agri Univ Ludhiana
- Stickel WH (1975) Some effects of pollutants in terristrial ecosystems. In: McIntyre AD, Mills CF (Eds) Ecologicant of the Interior Bureau of sport Fisheries and Wildlife. Patuxent Wildlife Research Centre
- Voss G, Sachssee K (1970) Red cell and plasma cholinesterase activities in microsamples of human and animal blood determined simultaneously by a modified acetylcholine/DTNB procedures. Toxicol Appl Pharmacol 16: 764-772

Umetsu N, Gross FH, Allahyari R, Abu-El-Haji S, Fukoto TR (1977) J of Agric Food Chem 25: 946-953 Wotton IDP (1964) Microanalysis Medical Biochemistry 4 edn Churchill J, Churchill A Ltd, London. Received February 28, 1988; accepted April 20, 1988.